

FIA Software 1RC API Documentation

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1 Module *fia_analysis*

FIA - Fluxomers Iterative Algorithm

SYNOPSIS:

```
fia_analysis {-i[---fia] PROJFILE | -t[---ftbl] PROJFILE} [-o[---output_file] OUTPUTFILE]
```

USAGE:

fia_analysis performs the following two main steps:

1) Metabolic pathway analysis.

Here FIA transforms the text file representing the metabolic network into mathematical matrices required later for the MFA optimization process.

2) Metabolic fluxes evaluation.

Here FIA tries to find the fluxes that best suit the given metabolic system. The output log of this stage is a file called `<PROJFILE>.out`, or else if specified by the "-o" flag.

fia_analysis supports two types of input file formats: FIA and 13CFlux FTBL.

The two are very much alike, except for the fact that the FIA format supports only declaration of uni-directional fluxes, and does not contain the FLUXES and INEQUALITIES sections.

For bi-directional fluxes, FIA simply defines two separated fluxes.

When analyzing 13CFlux FTBL files, *fia_analysis* looks for C=0 constraints in the FLUXES XCH section in order to determine directionality of fluxes.

OPTIONS:

`-i --fia`

Specifies that the metabolic pathway input file `<PROJFILE>` is a FIA formatted file. All fluxes are assumed unidirectional (for bi-directional specify 2 separated fluxes).

`-t --ftbl`

Specifies that the metabolic pathway input file `<PROJFILE>` is a 13CFlux FTBL formatted file. Flux is assumed unidirectional unless C=0 constraint is applied to it in the FLUXES XCH section. The FIA formatted file will be saved under `<PROJFILE>.fia`.

`-o --output_file`

optional.

Specifies the name of the result log output file. The default file name is `PROJFILE.out`

at least one of the above must be supplied.

USAGE EXAMPLE:

```
fia_analysis -t temp.ftbl
```

Runs analysis & evaluation of the network defined in the FTBL formatted input file `temp.ftbl`.

same as:

```
fia_analysis --ftbl temp.ftbl
```

```
fia_analysis --fia temp.fia -a |
```

Runs only network analysis on the given FIA formatted input file.

Same as:

```
fia_analysis -i temp.fia -a
fia_analysis -i temp.fia --analyze_only
fia_analysis --fia temp.fia --analyze_only
```

```
fia_analysis temp.fia -i -e
```

Runs only the fluxes evaluation on the FIA formatted input file temp.fia .

This assumes analysis has been done before on the temp.fia file.

Same as:

```
fia_analysis -i temp.fia --evaluate_only
fia_analysis --fia temp.fia -e
fia_analysis --fia temp.fia --evaluate_only
```

Version: 0.7.0

1.1 Functions

one_div_vec(*vec*)

This function returns a vector which is $1/\text{vec}$.

Parameters

vec: input vector
(*type=scipy.array*)

Return Value

$1/\text{vec}$ (element wise division).
(*type=scipy.array*)

sdiag(*a*)

Fast sparse diagonal matrix creator. Simply returns a sparse matrix with *a* as its diagonal.

Parameters

a: input array
(*type=array*)

Return Value

sparse matrix with *a* in its diagonal.
(*type=scipy.sparse matrix*)

expand_x_values(*cteq*)

This function transforms input with (\)'s into all the possible sets of inputs with 0's and 1's replacing the (\)s.

Parameters

cteq: Input string. (\)'s will be replaced with 1's and 0's.
(*type=string*)

Return Value

List of possible variations of cteq, with all the possible 0's and 1's replacements for \s in the input string.
(*type=list*)

create_ms_isotopomers_dict(*no_of_atoms, indices_to_change*)

This function returns all the possible permutations of no_of_atoms atoms, separated by the number of 1's in them. (*indices_to_change*) specifies which of the atoms can be changed (and the counting is then done only on these atoms). For example, if we call:

```
create_ms_isotopomers_dict(4, [0,1,2])
```

we get:

```
[['000x'], ['001x', '010x', '100x'], ['011x', '101x', '110x'], ['111x']]
```

The first element represents all 0's vec (for *indices_to_change*), the second a vector with only one 1 element, the third with two 1's elements, and the third with three 1's elements.

This function is used for mass-spectrometry measurements analysis.

Parameters

no_of_atoms: Number of atoms in the counted molecule.
(*type=number*)

indices_to_change: List of indices on which we want to count.
(*type=list*)

Return Value

List of lists - all possible permutations for the molecule, sorted by number of ones.
(*type=list*)

num2bin(*num*, *length*)

Converts number to binary format with at (*length*) bits.

Parameters

num: The number to be converted

(*type=number*)

length: Length of the output binary number.

(*type=number*)

Return Value

Binary string representing the input number. Zeros are left appended if necessary in order to maintain the required length.

(*type=string*)

save_text_vec(*filename*, *vec*)

This function saves the input vector in text TAB separated format. Used for MATLAB debugging purposes.

matlab_save_sparse_matrix(*filename*, *mat*)

This function saves the input matrix in text TAB separated format loadable by MATLAB. Used for MATLAB debugging purposes.

sort_dict_by_values(*d*)

This function sorts the input dictionary by its values.

null(*A*, *eps=1e-15*)

This function finds the null space of the incoming matrix *A*.

find_init_point(*self*)

This function generates the initial point given for the optimization process. It is based upon the standard init point finding methods of interior point algorithms: In order to find a valid solution for $S^*u = 0$, $U^* u \leq s$ we solve:

min(*s*)

s.t.:

$[I, -I]*u + [0*i, 10*i] \geq s$

$Su = S_b$

We then use this initial point with the propagation equation, resulting with the first point for the algorithm.

get_normG(*u, self*)

Main MFA optimization objective function value. This function calculates $\| \text{self.tG} * \mathbf{x} - \text{self.tB} \|$ for a given *u* vector by finding the valid *x* vector and then substituting it in the objective function.

get_normG_grad(*u, self*)

Main MFA optimization objective function gradient calculation. This function calculates $d(\| \text{self.tG} * \mathbf{x} - \text{self.tB} \|) / du$ for a given *u* vector.

load_data(*fname*)**tic**()**toc**()**main**(*argv*)

```

rand(d0, d1, dn, ...)
-----
Random values in a given shape.

Create an array of the given shape and propagate it with
random samples from a uniform distribution
over '[0, 1)'.

Parameters
-----
d0, d1, ..., dn : int
    Shape of the output.

Returns
-----
out : ndarray, shape '(d0, d1, ..., dn)'
    Random values.

See Also
-----
random

Notes
-----
This is a convenience function. If you want an interface that
takes a shape-tuple as the first argument, refer to
'random'.

Examples
-----
>>> np.random.rand(3,2)
array([[ 0.14022471,  0.96360618], #random
       [ 0.37601032,  0.25528411], #random
       [ 0.49313049,  0.94909878]]) #random

```

```

randn(d0, d1, dn, ...)
-----
Returns zero-mean, unit-variance Gaussian random numbers in an
array of shape (d0, d1, ..., dn).

Note: This is a convenience function. If you want an
      interface that takes a tuple as the first argument
      use numpy.random.standard_normal(shape_tuple).

```

1.2 Variables

Name	Description
<code>umfpack</code>	Value: <code>um.UmfpackContext()</code>
<code>prog_width</code>	Value: 40

continued on next page

Name	Description
display_prog	Value: False
ALLOW_THREADS	Value: 1
BUFSIZE	Value: 10000
CLIP	Value: 0
ERR_CALL	Value: 3
ERR_DEFAULT	Value: 0
ERR_DEFAULT2	Value: 2084
ERR_IGNORE	Value: 0
ERR_LOG	Value: 5
ERR_PRINT	Value: 4
ERR_RAISE	Value: 2
ERR_WARN	Value: 1
FLOATING_POINT_SUPPORT	Value: 1
FPE_DIVIDEBYZERO	Value: 1
FPE_INVALID	Value: 8
FPE_OVERFLOW	Value: 2
FPE_UNDERFLOW	Value: 4
False_	Value: False
Inf	Value: inf
Infinity	Value: inf
MAXDIMS	Value: 32
NAN	Value: nan
NINF	Value: -inf
NZERO	Value: -0.0
NaN	Value: nan
PINF	Value: inf
PZERO	Value: 0.0
RAISE	Value: 2
SHIFT_DIVIDEBYZERO	Value: 0
SHIFT_INVALID	Value: 9
SHIFT_OVERFLOW	Value: 3
SHIFT_UNDERFLOW	Value: 6
ScalarType	Value: (<type 'int'>, <type 'float'>, <type 'complex'>, <type 'l...
True_	Value: True
UFUNC_BUFSIZE_DEFAULT	Value: 10000
UFUNC_PYVALS_NAME	Value: 'UFUNC_PYVALS'
WRAP	Value: 1
__package__	Value: None
absolute	Value: <ufunc 'absolute'>
add	Value: <ufunc 'add'>
arccosh	Value: <ufunc 'arccosh'>
arcsinh	Value: <ufunc 'arcsinh'>
arctan	Value: <ufunc 'arctan'>
arctan2	Value: <ufunc 'arctan2'>
bitwise_and	Value: <ufunc 'bitwise_and'>
bitwise_not	Value: <ufunc 'invert'>
bitwise_or	Value: <ufunc 'bitwise_or'>
bitwise_xor	Value: <ufunc 'bitwise_xor'>

continued on next page

Name	Description
<code>c_</code>	Value: <numpy.lib.index_tricks.CClass object at 0x2bec6d0>
<code>cast</code>	Value: {<type 'numpy.int64'>: <function <lambda> at 0x2c68c80>, ...
<code>ceil</code>	Value: <ufunc 'ceil'>
<code>conj</code>	Value: <ufunc 'conjugate'>
<code>conjugate</code>	Value: <ufunc 'conjugate'>
<code>cos</code>	Value: <ufunc 'cos'>
<code>cosh</code>	Value: <ufunc 'cosh'>
<code>deg2rad</code>	Value: <ufunc 'deg2rad'>
<code>degrees</code>	Value: <ufunc 'degrees'>
<code>divide</code>	Value: <ufunc 'divide'>
<code>e</code>	Value: 2.71828182846
<code>equal</code>	Value: <ufunc 'equal'>
<code>exp</code>	Value: <ufunc 'exp'>
<code>exp2</code>	Value: <ufunc 'exp2'>
<code>expm1</code>	Value: <ufunc 'expm1'>
<code>fabs</code>	Value: <ufunc 'fabs'>
<code>floor</code>	Value: <ufunc 'floor'>
<code>floor_divide</code>	Value: <ufunc 'floor_divide'>
<code>fmax</code>	Value: <ufunc 'fmax'>
<code>fmin</code>	Value: <ufunc 'fmin'>
<code>fmod</code>	Value: <ufunc 'fmod'>
<code>frexp</code>	Value: <ufunc 'frexp'>
<code>greater</code>	Value: <ufunc 'greater'>
<code>greater_equal</code>	Value: <ufunc 'greater_equal'>
<code>hypot</code>	Value: <ufunc 'hypot'>
<code>index_exp</code>	Value: <numpy.lib.index_tricks.IndexExpression object at 0x2bec790>
<code>inf</code>	Value: inf
<code>infty</code>	Value: inf
<code>invert</code>	Value: <ufunc 'invert'>
<code>isfinite</code>	Value: <ufunc 'isfinite'>
<code>isinf</code>	Value: <ufunc 'isinf'>
<code>isnan</code>	Value: <ufunc 'isnan'>
<code>ldexp</code>	Value: <ufunc 'ldexp'>
<code>left_shift</code>	Value: <ufunc 'left_shift'>
<code>less</code>	Value: <ufunc 'less'>
<code>less_equal</code>	Value: <ufunc 'less_equal'>
<code>little_endian</code>	Value: True
<code>log1p</code>	Value: <ufunc 'log1p'>
<code>logaddexp</code>	Value: <ufunc 'logaddexp'>
<code>logaddexp2</code>	Value: <ufunc 'logaddexp2'>
<code>logical_and</code>	Value: <ufunc 'logical_and'>
<code>logical_not</code>	Value: <ufunc 'logical_not'>
<code>logical_or</code>	Value: <ufunc 'logical_or'>
<code>logical_xor</code>	Value: <ufunc 'logical_xor'>
<code>maximum</code>	Value: <ufunc 'maximum'>
<code>mgrid</code>	Value: <numpy.lib.index_tricks.nd_grid object at 0x2bec510>

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Name	Description
minimum	Value: <ufunc 'minimum'>
mod	Value: <ufunc 'remainder'>
modf	Value: <ufunc 'modf'>
multiply	Value: <ufunc 'multiply'>
nan	Value: nan
nbytes	Value: {<type 'numpy.int64'>: 8, <type 'numpy.int16'>: 2, <type ...
negative	Value: <ufunc 'negative'>
newaxis	Value: None
not_equal	Value: <ufunc 'not_equal'>
ogrid	Value: <numpy.lib.index_tricks.nd_grid object at 0x2bec550>
ones_like	Value: <ufunc 'ones_like'>
pi	Value: 3.14159265359
r_	Value: <numpy.lib.index_tricks.RClass object at 0x2bec610>
rad2deg	Value: <ufunc 'rad2deg'>
radians	Value: <ufunc 'radians'>
reciprocal	Value: <ufunc 'reciprocal'>
remainder	Value: <ufunc 'remainder'>
right_shift	Value: <ufunc 'right_shift'>
rint	Value: <ufunc 'rint'>
s_	Value: <numpy.lib.index_tricks.IndexExpression object at 0x2bec810>
sctypeDict	Value: {0: <type 'numpy.bool_'>, 1: <type 'numpy.int8'>, 2: <typ...
sctypeNA	Value: {'?': 'Bool', 'B': 'UInt8', 'Bool': <type 'numpy.bool_'>, ...
sctypes	Value: {'complex': [<type 'numpy.complex64'>, <type 'numpy.compl...
sign	Value: <ufunc 'sign'>
signbit	Value: <ufunc 'signbit'>
sin	Value: <ufunc 'sin'>
sinh	Value: <ufunc 'sinh'>
square	Value: <ufunc 'square'>
subtract	Value: <ufunc 'subtract'>
tan	Value: <ufunc 'tan'>
tanh	Value: <ufunc 'tanh'>
true_divide	Value: <ufunc 'true_divide'>
trunc	Value: <ufunc 'trunc'>
typeDict	Value: {0: <type 'numpy.bool_'>, 1: <type 'numpy.int8'>, 2: <typ...
typeNA	Value: {'?': 'Bool', 'B': 'UInt8', 'Bool': <type 'numpy.bool_'>, ...
typecodes	Value: {'All': '?bhilqpBHILQPfdgFDGSUV0', 'AllFloat': 'fdgFDG', ...

1.3 Class MyWriter

A file-handler replacement that duplicates the actions applied to the handler to a log file handler as well.

1.3.1 Methods

<code>__init__(self, stdout, filename)</code>
Initialization of the file-handler duplicator.
Parameters
stdout: Original file handler to be forked (<i>type=file</i>)
filename: Name of the log file for which actions should be replicated into (<i>type=string</i>)

<code>write(self, text)</code>
Replacement for the file handler's write method.

<code>flush(self)</code>
Replacement for the file handler's flush method.

<code>close(self)</code>
Replacement for the file handler's close method.

1.4 Class FtblFile

This class is the main object of the FIA algorithm. Its main purposes are:

1. Conversion of FIA and FTBL files into python mathematical objects.
2. Solution of the MFA optimization problem

1.4.1 Methods

<code>__init__(self, projname, inpfiletype)</code>
Parameters
projname: Name of the project to be analyzed. (<i>type=string</i>)
inpfiletype: Type of the input file to be analyzed. Can be either 'fia' (for fia input files) or 'ftbl' (for ftbl input files) (<i>type=string</i>)

save_ftblfile(*self*, *filename*)

This function saves the entire FtblFile class to a file using python's pickle package. This enables the user to use the data analyzed later without the need of recalculating the transition or LU matrices (which can take a bit of a time).

save_data(*self*)

MATLAB debugging function. This function saves the mathematical objects used for the optimization process in files in order to be analyzed by MATLAB^a.

^a<http://www.mathworks.com>

get_flux_name(*self*, *flux_num*)

returns the name of fluxer whose index is flux_num.

Parameters

flux_num: The number of the fluxomer we are looking for.
(*type=number*)

Return Value

The name of the fluxomer.
(*type=string*)

load_file(*self*, *filename*)

This function loads the input file into the FtblClass object by:

1. Dropping comments (lines starting with "//")
2. Replacing EOL "\n" or "\r" with ""
3. Erasing spaces

Eventually we are left with a file containing only TABs. This object is then transformed into an array of lists - every array element is a line of the parsed file, and splitted by TABs into a list object.

Parameters

filename: Name of the input file to be examined.
(*type=string*)

Return Value

List of lists containing the TAB splitted parsed version of the input file.
(*type=list*)

find_segment(*self*, *seg_name*)

This function finds the block "seg_name" block segment in the parsed input file `self.ftbl_file` object. New segments begin with every non-idented line in the input file. Their names are the first string in these lines.

Parameters

seg_name: Name of the segment to be analyzed
(*type=string*)

Return Value

A list containing the segment's rows, tab separated.
(*type=list*)

create_pools_dict(*self*)

This function creates the initial metabolic pools dictionary object by analyzing the NETWORK section of the input file. The function creates the `self.pools` dictionary object - Metabolic pools dictionary for which the keys are pools names, and the values are the number of carbon atoms for each pool.

create_flux_dict(*self*)

This function creates and updates the main building blocks objects used for the metabolic pathway analysis.

(section) Classification of fluxes

Classification of fluxes into: uni-directional fluxes, bi-directional fluxes (FTBL files only) and input/output fluxes. The uni and bi directional analysis is only needed for FTBL files (FIA assumes all the fluxes are uni-directional). The decision is made by looking for 0 constrained exchange fluxes in the FLUXES section of the FTBL file.

(section) Re-generation of the NETWORK section - adding necessary virtual fluxes

When we are given with a chemical reaction that transforms two elements of the same metabolic pool into others, we translate it into a regular multi-input chemical reaction by adding a virtual pool and flux to our system:

As an example:

`f: A + A -> B + C` (flux "f" transforms A+A into B+C)

is transformed into:

`f: A + A_s -> B + C` (flux "f" transforms A+A_s into B+C)
`f_A: A -> A_s` (flux "f_A" transforms A into A_s)

This transformation is being done on the input file object itself (hence by changing the parsed NETWORK segment of the input file). The same applies for reversed fluxes of bi-directional FTBL files fluxes.

(section) Analysis of extra-cellular pools

Finding pools which are global input or output to the system, and save them in the `self.excell_pools` object. These pools are recognized as pools with no input / output fluxes going into / exiting them.

(section) Re-generation of the NETWORK section - adding necessary reversed fluxes

The mathematical analysis of the problem consists of flux vector representing uni-directional fluxes. Due to this reason, we need to transform the NETWORK section to contain only uni-directional fluxes by adding to it necessary reversed fluxes. This only applies when FTBL input files are supplied (since again, FIA files are assumed to contain only uni-directional fluxes in anyway).

(section) Generation of the global fluxes vector

Generation of the global fluxes (as opposed to fluxomers) vector dictionary object. The dictionary `self.flux_nw_vec` keys are the names of the global fluxes in our system, and the values are the number of carbon atoms each of them transforms.

(section) Generation of the equality matrix

The equality matrix `self.S` and its associated equality vector `self.S_b` are the mathematical constraints of the global optimization problem that should apply on the global fluxes vector (hence `self.S * x == self.S_b`). The equality matrix is constructed out of 2 main elements:

1. The stoichiometric equations - as analyzed from the parsed NETWORK section of the input file
2. The equalities constraints from the EQUALITIES section of the input file.

(section) Generation of the equality measurements

Equalities arise from the constraints section of the FLUXES section (only applies to FTBL

get_metabolic_flux_index(*self*, *flux_name*)

This function returns the index of the global metabolic flux "flux_name".

Parameters

flux_name: The name of the flux we want to find.
(*type=string*)

Return Value

The index of the global metabolic flux "flux_name".
(*type=number*)

create_flux_meas_mat(*self*)

This function parses the FLUX_MEASUREMENTS segment of the ftbl file. It creates the following matrices:

1. **self.flux_meas_mat** - The matrix choosing the measured metabolic fluxes out of the fluxomer vector.
2. **self.flux_meas_values** - The measured values.
3. **self.flux_meas_variance** - The measurement variance vector.

Eventually we will search for:

$$M\{\min [(\mathbf{self.flux_meas_mat} * \mathbf{x} - \mathbf{self.flux_meas_values}) .T * \mathbf{diag}(\mathbf{self.flux_meas_variance}) * (\mathbf{self.flux_meas_mat} * \mathbf{x} - \mathbf{self.flux_meas_values})]\}$$

create_global_meas_mat(*self*)

This function creates the objective function for the MFA optimization problem (the global "G" measurement matrix).

The objective of the MFA problem is to minimize:

$$M\{\|\mathbf{tG} * \mathbf{x} - \mathbf{tB}\|^2\}$$

The matrix **self.tG** is created by concatenation of the 3 possible measurements matrices:

1. Label measurements (**self.labelinp_num** and **self.labelinp_denom**).
2. Fluxes measurement (**self.labelmeas_num** and **self.labelmeas_denom**).
3. Mass-Spectrometry measurements (**self.ms_meas_num** and **self.ms_meas_denom**).

The vector **self.tB** is created by concatenation of appropriate values of the above.

trnasform_eq_into_matrix(*self*, *eq*)

This function transforms string equation of fluxes into a fluxomers stoichiometric matrix. For example:

```
"*+flux1/0*-flux2/1" -> [1 0 0 0; 0 0 0 -1])
```

Parameters

eq: An string equation. Every equation element is seperated by either "*" or "-" with the fluxomer name following it. No preciding numbers are allowed, only plain fluxomer names. For multiple instances of the same fluxomer, simply add/substract it more than once to the equation.

(*type=string*)

Return Value

The fluxomer stoichiometric matrix represented by eq.

(*type=CSR matrix*)

create_label_meas_eq(*self*)

This function parses the LABEL_MEASUREMENTS segment of the ftbl file. The output of the function is the matrices and vectors:

1. `self.labelmeas_num` - The label measurements numerator matrix
2. `self.labelmeas_denom` - The label measurements denominator matrix
3. `self.labelmeas_value` - The label measurements values vector
4. `self.labelmeas_var` - The label measurements values vector

The objective function will be to minimize:

$$M\{ \|\text{diag}(\text{self.labelmeas_var}) * [\text{self.labelmeas_num} - \text{diag}(\text{self.labelmeas_value}) * \text{self.labelmeas_denom}]\|$$
create_ms_meas_eq(*self*)

This function parses the MASS_SPECTROMETRY segment of the ftbl file. The output of the function is the matrices and vectors:

1. `self.ms_meas_num` - The MS measurements numerator matrix
2. `self.ms_meas_denom` - The MS measurements denominator matrix
3. `self.ms_meas_value` - The MS measurements values vector
4. `self.ms_meas_var` - The MS measurements values vector

We always treat these measurement as a ratio measurement between the MS values, relative to the 0 MS measurement (hence we add to the objective the ratios: `mass1/mass0`, `mass2/mass0` etc').

The objective function will be to minimize:

$$M\{ \|\text{diag}(\text{self.ms_meas_var}) * [\text{self.ms_meas_num} - \text{diag}(\text{self.ms_meas_value}) * \text{self.ms_meas_denom}]\|$$

create_U_matrix(self)

This function constructs the U matrix, which transforms the fluxomers vector into the metaoblic fluxes vector. hence:

$$M\{u = U*x\}$$

where:

u - the metabolic flux vector
x - the fluxomers vector

create_inp_eq_matrix(self)

This function parses the LABEL_INPUT segment. The output of the function is the matrices and vectors:

1. `self.labelinp_num` - The flux value measurements numerator matrix
2. `self.labelinp_denom` - The flux value measurements denominator matrix
3. `self.labelinp_value` - The flux value measurements values vector

The objective function will be to minimize:

$$M\{ ||diag(self.labelinp_var) * [self.labelinp_num - diag(self.labelinp_value)] * self.labelinp_denom || \}$$

Note: since not supplied by the FTBL files, the value of `self.labelinp_value` is a global assigned constant.

create_label_transition_mat(self)

This function creates the propogation transition matrix:

$$\backslash M\{x(t) = P x(t-1)\} .$$

P is provided using the following sub matrices:

1. `self.trans_mat_a` - the H_1 matrix (in CSR format).
2. `self.trans_mat_b` - the H_2 matrix (in CSR format).
3. `self.trans_mat_a_gamma` - the g_1 matrix (in CSR format).
4. `self.trans_mat_a_beta` - the g_2 matrix (in CSR format).
5. `self.trans_mat_b_beta` - the g_3 matrix (in CSR format)

In addition, this function creates the beta (ratios) vector:

1. `self.beta_vec` - the text names of the beta vector elements
2. `self.beta_vec_M` - the numerator for construction of the beta vec.
3. `self.beta_vec_N` - the denominator for constuction of the beta vec.

eventually, we have:

$$M\{beta_vec = (self.beta_vec_M * u) / (self.beta_vec_N * u)\}$$

create_lu_trans_matrix(*self*)

This function computes the LU decomposition of the propogation transition matrices. We find a factorization for:

$$L\{\text{self.trans_mat_a}\} \text{ and } L\{\text{self.trans_mat_b}\}$$

by finding the LU factorizaiton of the matix constructed out of concatanation of the two above.

The result are the matrices:

1. **self.a11** - the left side of the pseudo LU of self.trans_mat_a
2. **self.a12** - the left side of the pseudo LU of self.trans_mat_b
3. **self.au** - the right side of the pseudo LU factorizaiton of both self.trans_mat_a and self.trans_mat_b .

evaluate(*self*)

This function runs the main optimization process. It uses the `scipy.optimize.fmin_slsqp` for the actual optimization process, and prints out the results in a TABed formatted table.

1.4.2 Class Variables

Name	Description
filename	Value: ''

1.4.3 Instance Variables

Name	Description
ProjName	Project's name
labelinp_var	Label input variance (it is not supplied with FTBL files).
InputFileType	input file type (fia or ftbl)
excell_pools	Dictionary of the external (global input or output) metabolic pools
ftbl_file	Parsed input file object (TAB spliited lists for the various lines of the input file) Value: ''
pools	Metabolic pools dictionary (Keys are pools names, values are the number of carbon atoms for each pool)

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